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(FILE 'HOME' ENTERED AT 15:02:39 ON 29 JAN 2003)

FILE 'BIOSIS, CAPLUS, EMBASE, MEDLINE, CANCERLIT' ENTERED AT 15:02:57 ON
29 JAN 2003

L1 183 S APTAMER AND CONCENTR?
L2 83 DUPLICATE REMOVE L1 (100 DUPLICATES REMOVED)
L3 23 S L2 AND NUCLEIC ACID

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(FILE 'HOME' ENTERED AT 12:51:16 ON 29 JAN 2003)

FILE 'BIOSIS, CAPLUS, EMBASE, MEDLINE, CANCERLIT' ENTERED AT 12:52:15 ON
29 JAN 2003

L1	121 S APTAMER AND QUANTI?
L2	46 S L1 AND NUCLEIC ACID
L3	33 DUPLICATE REMOVE L2 (13 DUPLICATES REMOVED)
L4	40 S APTAMER AND AMOUNT
L5	10 S L4 AND NUCLEIC ACID
L6	60 DUPLICATE REMOVE L1 (61 DUPLICATES REMOVED)

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(FILE 'HOME' ENTERED AT 12:51:16 ON 29 JAN 2003)

FILE 'BIOSIS, CAPLUS, EMBASE, MEDLINE, CANCERLIT' ENTERED AT 12:52:15 ON
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L1 183 S APTAMER AND CONCENTR?
L2 83 DUPLICATE REMOVE L1 (100 DUPLICATES REMOVED)
L3 23 S L2 AND NUCLEIC ACID

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(FILE 'HOME' ENTERED AT 11:13:26 ON 29 JAN 2003)

FILE 'BIOSIS, CAPLUS, EMBASE, MEDLINE, CANCERLIT' ENTERED AT 11:13:50 ON
29 JAN 2003

L1	2389 S APTAMER
L2	121 S L1 AND QUANTI?
L3	46 S L2 AND NUCLEIC ACID
L4	33 DUPLICATE REMOVE L3 (13 DUPLICATES REMOVED)

L5 ANSWER 6 OF 10 CAPLUS COPYRIGHT 2003 ACS
 AN 2000:299876 CAPLUS
 DN 133:204835
 TI **Aptamers**: another use for oligonucleotides
 AU Conrad, Richard C.
 CS Lilly Research Labs, Lilly Corporate Center, Eli Lilly and Co.,
 Indianapolis, IN, 46285, USA
 SO Antisense Technology in the Central Nervous System (1999), 195-217.
 Editor(s): Leslie, Ronald A.; Hunter, A. Jackie; Robertson, Harold A.
 Publisher: Oxford University Press, Oxford, UK.
 CODEN: 68XUAZ
 DT Conference; General Review
 LA English
 CC 9-0 (Biochemical Methods)
 AB A review with 32 refs. With the advent of simple means for replication of
nucleic acids in vitro and the ability to chem.
 synthesize large **amts.** of **nucleic acids** in
 the 100 nucleotide (nt) range, novel **nucleic acids** with
 previously unknown binding functionalities can be created and propagated
 in vitro. Variations of these and other techniques can be used to find
 these novel binding **nucleic acids**, using a procedure
 of cyclized amplification and selection steps referred to as SELEX
 (systematic evolution of ligands by exponential enrichment). The actual
nucleic acid ligands are called **aptamers**.
 This chapter is meant to serve as a primer for the selection of
aptamers, providing a basic approach to SELEX.
 ST review SELEX **nucleic acid aptamer** selection
 IT RNA
 RL: BSU (Biological study, unclassified); BIOL (Biological study)
 (**aptamers**; selection of **aptamers** using SELEX)
 IT **Nucleic acids**
 RL: BSU (Biological study, unclassified); BIOL (Biological study)
 (ligands, **aptamers**; selection of **aptamers** using
 SELEX)
 RE.CNT 32 THERE ARE 32 CITED REFERENCES AVAILABLE FOR THIS RECORD
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L5 ANSWER 7 OF 10 MEDLINE

L6 ANSWER 51 OF 60 CAPLUS COPYRIGHT 2003 ACS DUPLICATE 19
AN 2000:72134 CAPLUS
DN 133:55582

TI The use of **aptamers** in large arrays for molecular diagnostics
AU Brody, Edward N.; Willis, Michael C.; Smith, Jonathan D.; Jayasena,
Sumedha; Zichi, Dominic; Gold, Larry

CS NeXstar Pharmaceuticals, Inc, Boulder, CO, 80301, USA

SO Molecular Diagnosis (1999), 4(4), 381-388

CODEN: MDIAFU; ISSN: 1084-8592

PB Churchill Livingstone

DT Journal

LA English

CC 9-16 (Biochemical Methods)

Section cross-reference(s): 3, 6

AB Background: **Aptamers** are single-stranded oligonucleotides derived from an in vitro evolution protocol called systematic evolution of ligands by exponential enrichment (SELEX). They bind tightly and specifically to target mols.; most **aptamers** to proteins bind with Kds (equil. dissocn. const.) in the range of 1 pM to 1 nM. Methods and Results: The SELEX protocol has been automated; therefore, hundreds to thousands of **aptamers** can be made in an economically feasible fashion. Blood and urine can be analyzed on chips that capture and **quantitate** proteins. SELEX has been adapted to the use of 5-bromo (5-Br) and 5-iodo (5-I) deoxyuridine residues. These halogenated bases can be specifically cross-linked to proteins. Selection pressure during in vitro evolution can be applied for both binding specificity and specific photo-cross-linkability. These are sufficiently independent parameters to allow one reagent, a photo-cross-linkable **aptamer**, to substitute for two reagents, the capture antibody and the detection antibody, in a typical sandwich array. After a cycle of binding, washing, crosslinking, and detergent washing, proteins will be specifically and covalently linked to their cognate **aptamers**. Conclusions: Because no other proteins are present on the chips, protein-specific stain will now show a meaningful array of pixels on the chip. Learning algorithms and retrospective studies should lead to a robust, simple, diagnostic chip.

ST **aptamer** oligonucleotide mol diagnosis protein analysis method

IT Oligonucleotides

RL: ARG (Analytical reagent use); ARU (Analytical role, unclassified); BPR (Biological process); BSU (Biological study, unclassified); ANST (Analytical study); BIOL (Biological study); PROC (Process); USES (Uses) (SELEX **aptamers**; use of **aptamers** in large arrays for mol. diagnostics)

IT Diagnosis

(agents; use of **aptamers** in large arrays for mol. diagnostics)

IT Diagnosis

(mol.; use of **aptamers** in large arrays for mol. diagnostics)

IT Proteins, general, analysis

RL: ANT (Analyte); BPR (Biological process); BSU (Biological study, unclassified); ANST (Analytical study); BIOL (Biological study); PROC (Process)

(use of **aptamers** in large arrays for mol. diagnostics)

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